

## Antioxidant Capacity and Tentative Identification of Polyphenolic Compounds of Cones of Selected Coniferous Species

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**Abstract** – The cones of coniferous species are a waste biomass byproduct that can be potentially utilized for a variety of purposes. One of the many application fields is the extraction of bioactive materials, particularly antioxidant polyphenols. Scientific literature on the antioxidant content of coniferous cones at different ripening stages is limited. In this study, we conducted a comparative analysis of the antioxidant content of selected taxa that are either common in Hungary or that have not yet been investigated in the scientific literature in any great detail (*Cedrus atlantica*, *Larix decidua*, *Picea abies*, *Pinus mugo*, *Pinus nigra*, *Pinus sylvestris*, *Pinus wallichiana*, *Tsuga canadensis*, *Tsuga heterophylla*, *Chamaecyparis lawsoniana*, *Taxodium distichum*, *Thuja occidentalis*, *Metasequoia glyptostroboides*, *Thuja orientalis*, *Cryptomeria japonica*, *Cunninghamia lanceolata*). A comparison of green, mature and opened cones was performed for the assigned taxa. Folin-Ciocalteu total polyphenol content (TPC), ferric reducing antioxidant power (FRAP) and 2,2-diphenyl-1-picrylhydrazyl (DPPH) assays were used to assess the antioxidant contents. Overall antioxidant power was determined by a scoring system that combined the three assay results. In general, best values were found for green cones, followed by mature, and opened cones for each taxon. *Tsuga canadensis*, *Metasequoia glyptostroboides*, *Chamaecyparis lawsoniana*, *Cryptomeria japonica*, *Thuja orientalis* and *Picea abies* all contained high amounts of antioxidants in both green and mature cones and attained the highest scores. High-performance liquid chromatographic/tandem mass spectrometric profiling of the cone polyphenols was also completed for selected samples. Results provide a basis for future bioactivity testing of these samples.

**coniferous species / cones / antioxidants / HPLC-MS/MS**

**Kivonat** – Túlevelű taxonok tobozainak antioxidáns kapacitása és polifenolos vegyületeinek vizsgálata. A túlevelű fajok tobozai olyan hulladék biomasszát képviselnek, melyeket többféle célra is lehetne használni. Az egyik ilyen felhasználási terület a bioaktív anyagok, például antioxidáns polifenolok kinyerése. A tobozérés különböző fenofázisaiban az antioxidáns tartalomra vonatkozó szakirodalmi adatok hiányosak. Jelen cikkben olyan taxonok vizsgálatát végeztük el, melyek vagy Magyarországon gyakoriak, vagy még nem történt meg a vizsgálatuk (*Cedrus atlantica*, *Larix decidua*, *Picea abies*, *Pinus mugo*, *Pinus nigra*, *Pinus sylvestris*, *Pinus wallichiana*, *Tsuga canadensis*, *Tsuga heterophylla*, *Chamaecyparis lawsoniana*, *Taxodium distichum*, *Thuja occidentalis*, *Metasequoia glyptostroboides*, *Thuja orientalis*, *Cryptomeria japonica*, *Cunninghamia lanceolata*). Elvégeztük a zöld, érett és lehullott tobozok összehasonlító vizsgálatát az összes polifenol tartalom (Folin-Ciocalteu), a FRAP (ferric reducing antioxidant power) és a DPPH (2,2-diphenyl-1-picrylhydrazyl) antioxidáns kapacitás meghatározási módszerek segítségével. Az összesített antioxidáns kapacitás kiértékelése a

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három módszer egyesítésével, egy pontrendszer segítségével történt meg. Összességében a legnagyobb antioxidáns kapacitást a zöld tobozokra mértük, a legalacsonyabbat a lehullott tobozokra mindegyik taxon esetében. A legmagasabb pontszámot a *Tsuga canadensis*, *Metasequoia glyptostroboides*, *Chamaecyparis lawsoniana*, *Cryptomeria japonica*, *Thuja orientalis* és *Picea abies* zöld és értett tobozai kapták. A kiválasztott minták esetében elvégeztük a polifenol készlet profilozását nagyhatékonyságú folyadékkromatográfiás/tandem tömegspektrometriás eljárással. Az eredmények alapját képezhetik ezen minták bioaktivitás-vizsgálatának.

## tűlevelű fajok / toboz / antioxidánsok / HPLC-MS/MS

### 1 INTRODUCTION

Forestry, logging and timber production wastes (e.g. leaves, wood bark, cones, etc.) can be a rich source of antioxidant compounds (Dedrie et al. 2015, Bouras et al. 2016) with potential utilization fields including the production of healthcare-related products (Packer et al. 1999, Dzialo et al. 2016, Watson et al. 2018), natural food preservatives and ingredients (Coté et al. 2011, Gyawali – Ibrahim 2014, Kobus-Cisowska et al. 2014, Frydman et al. 2005), natural growth bioregulators (Popa et al. 2008, Vyvyan 2002) as well as silver nanoparticles (Fahimirada et al. 2019, Rolim et al. 2019) to name but a few.

As waste biomass basic materials, cones represent a biomass exclusively born by coniferous trees and shrubs belonging to one of the over 615 living species (Auders – Spicer 2012). Conifers bear “seed-cones” and “pollen-cones” out of which the female seed-cones are simply referred to as “cones”; these were the exclusive subject of the present study.

The primary use of forest tree cones has been seed extraction for the production of forestry propagation material. In the Mediterranean region the edible seeds of stone pine cones (*Pinus pinea* L.) are one of the most important tree nuts (Kemerli-Kalbaran – Ozdemir 2019). The empty cones are usually burned (Aniszewska – Bereza 2014) in an uncompressed state or can be converted to briquettes (Gendek et al. 2018). The cones *Juniperus* spp. have traditionally been used for flavouring purposes (Lesjak et al. 2011), while the cone extracts and essential oils of *Pinus*, *Thuja*, and *Cedrus* spp. have been used by traditional medicine for various beneficial (e.g. anti-inflammatory, antioxidant, antiseptic, antifungal, antimicrobial, analgesic etc.) health effects (Watanabe et al. 1995, Lesjak et al. 2011, Süntar et al. 2012, Djouahri et al. 2014). The cone extracts of *Pinus parviflora* Siebold et Zucc. were shown to be very powerful against HIV and influenza viruses (Nagata et al. 1990) and were also shown to possess significant antimutagenic and anticancer effects (Nagasawa et al. 1992). The cone and essential oil extracts of *Metasequoia glyptostroboides* (Bajpai et al. 2014), *Juniperus sibirica* Burgsdorf. (Lesjak et al., 2011), *Tetraclinis articulata* (Vahl) Mast. (Djouahri et al. 2014), *Cupressus sempervirens* var. *pyramidalis* (L.) (Tumen et al. 2012) and of *Pinus* spp. (Süntar et al. 2012, Bradley et al. 2014, Tümen et al. 2018, Wang et al. 2019) were recently shown to have significant beneficial effects on human health. The latest results indicate that pine cone and pine cone extracts can be used for their various useful properties, e.g. being a source as dietary fibre (Kartal – Ozturk 2016), or starting materials for the production of coagulants (Hussain et al. 2019) and adsorbents (Kupeta et al. 2018, Mtshatsheni et al. 2019).

Despite the listed results, the literature lacks systematic research of the antioxidant composition of cones and the assessment of their role as a source of natural antioxidants. Moreover, sample collection times in the presented examples – more specifically, the phenophase of cone maturity – have rarely been documented in the literature. Recently Hofmann et al. (2020) concluded a systematic research using optimized extraction conditions and multiassay evaluation for the assessment of the antioxidant content of coniferous cones while respecting the phenophase of cone maturity; however, this study included only 6 taxa.

The aim of the present research was to extend previous studies (Hofmann et al. 2020) by investigating altogether 16 taxa including Atlas cedar (*Cedrus atlantica* Endl.), European larch (*Larix decidua* Mill.), Norway spruce (*Picea abies* H. Karst.), mountain pine (*Pinus mugo* Turra), black pine (*Pinus nigra* J.F. Arnold), Scots pine (*Pinus sylvestris* L.), Himalayan pine (*Pinus wallichiana* A. B. Jacks.), eastern hemlock (*Tsuga canadensis* (L.) Carrière), western hemlock (*Tsuga heterophylla* (Raf.) Sarg.), Lawson cypress (*Chamaecyparis lawsoniana* (A. Murray) Parl.), bald cypress (*Taxodium distichum* (L.) Rich.), northern white-cedar (*Thuja occidentalis* L.), dawn redwood (*Metasequoia glyptostroboides* Hu and W. C. Cheng), Chinese arborvitae (*Thuja orientalis* L.), Japanese cedar (*Cryptomeria japonica* (L.f.) D. Don) and China fir (*Cunninghamia lanceolata* (Lamb.) Hook).

Antioxidant properties were assayed by the Folin-Ciocalteu total polyphenol content (TPC), ferric reducing antioxidant power (FRAP) and 2,2-diphenyl-1-picrylhydrazyl (DPPH) methods. The evaluation of the overall antioxidant power was accomplished by a scoring system, which combined the results of the TPC, FRAP, and DPPH methods. In this manner a comprehensive evaluation of the results between various samples with potentially different antioxidant compositions was achieved.

The polyphenol profile of most relevant samples with the highest antioxidant potential was also investigated using high-performance liquid chromatography/multistage mass spectrometry (HPLC-MS/MS) in order to identify the structure of major antioxidant compounds, primarily polyphenols.

## 2 MATERIALS AND METHODS

### 2.1 Chemicals and reagents

Double distilled water was prepared for the extractions using conventional distillation equipment. LCMS grade acetonitrile, and acetone was obtained from VWR International (Budapest, Hungary). Gallic acid, ascorbic acid, DPPH, 2,4,6-tripyridyl-S-triazine (TPTZ), iron(III)-chloride, acetic acid, sodium acetate, hydrochloric acid, and sodium carbonate were obtained from Sigma-Aldrich (Budapest, Hungary). Folin-Ciocalteu reagent was purchased from Merck (Darmstadt, Germany).

### 2.2 Sample collection and extraction

Sample collection occurred at the Botanical Garden of the University of Sopron in Sopron, Hungary between July-October 2019. Three ripening stages were sampled: green cones (collected in July when cones are green, yet nearly at their full size at the final year of maturation), mature cones (collected in August/September when the cones turned brown in colour and scales began to open) and opened cones (taken in September/October, at a fully opened state having released their seeds and found on trees or to the ground). One healthy individual of each taxon was sampled by collecting a minimum of 10 cones from different parts of the crown at each sampling occasion. Cone samples were put into sealed plastic bags and stored at -20°C until processing. Prior to extraction, samples were thawed and ground. Ultrasonic extraction was performed using an Elma Transsonic T570 ultrasonic bath (Elma Schmidbauer GmbH, Singen, Germany) as follows: 0.45 g ground sample was homogenized with 45 ml acetone:water 80:20 v/v in a 50 ml centrifuge tube and sonicated for 3 x 10 min as described by Hofmann et al. (2020). One extraction was conducted for each sample.

## 2.3 Determination of antioxidant properties

TPC, FRAP, and DPPH measurements were run in triplicates using of a Hitachi U-1500 type spectrophotometer (Hitachi Ltd., Tokyo, Japan).

### 2.3.1 Total polyphenol content (TPC)

TPC determination was completed by applying the Folin-Ciocalteu assay (Singleton – Rossi 1965) using gallic acid as the standard: 0.5 ml extract solution was mixed with 2.5 ml 10-fold diluted Folin-Ciocalteu reagent. After 1 min, 2 ml 0.7 M Na<sub>2</sub>CO<sub>3</sub> solution was added and the reaction mixture was heated for 5 min in a 50 °C water bath. Reaction was stopped by cooling to room temperature in a cold water bath. Solution absorbance was measured at 760 nm. The results were expressed as mg equivalents of gallic acid/g dry bark units (mg GAE/g d.w.).

### 2.3.2 FRAP antioxidant capacity

The method described by Benzie – Strain (1996) was applied for the measurement of the FRAP antioxidant capacity at 593 nm using ascorbic acid as a standard. FRAP reagent was prepared as follows: 25 ml of 10 mM TPTZ solution (aqueous with 84 µl cc. HCl) was mixed with 250 ml of acetate buffer (300 mM, pH 3.6) and 25 mL of 20 mM aqueous FeCl<sub>3</sub> solution. Fifty µl sample was mixed with 1500 µl FRAP reagent in glass test tubes at ambient temperature and after 5 min reaction time absorbance was measured. Results were given in mg equivalents of ascorbic acid/g dry weight (mg AAE/g dw.).

### 2.3.3 DPPH antioxidant capacity

The slightly modified method of Sharma – Bhat (2009) was used for running the DPPH assay as follows: 2090 µl unbuffered methanol was mixed with 900 µl 2×10<sup>-4</sup> M methanolic DPPH solution and 10 µl extract. After 30 min incubation at room temperature in the dark, the decrease in absorbance was determined at 515 nm. Results were calculated in IC<sub>50</sub> (50% inhibition concentration) values in µg extractives/ml assay (µg/ml) units, representing the amount of extractives which will react with 50% of the added DPPH• radicals in the total assay volume (3 ml).

## 2.4 HPLC-MS/MS analyses

Separation of the cone extracts of Norway spruce and eastern hemlock was achieved using a Shimadzu LC-20 type high-performance liquid chromatograph coupled with a Shimadzu SPD-M20A type diode array detector (PDA) (Shimadzu Corporation, Kyoto, Japan) and an AB Sciex 3200 QTrap triple quadrupole/linear ion trap mass spectrometric (MS) detector (AB Sciex, Framingham, USA). A Phenomenex Synergy Fusion-RP 80A, 250 mm x 4.6 mm, 4µm column was used for the separation with a Phenomenex SecurityGuard ULTRA LC type guard column (Phenomenex Inc., Torrance, USA) at 40°C. The injection volume was 15 µl. The binary gradient of A (H<sub>2</sub>O + 0.1% HCOOH) and B (CH<sub>3</sub>CN + 0.1% HCOOH) solvents was run with 1.2 ml/min flow-rate using the following time gradient: 3% B (0-4 min), 6% B (10 min), 20% B (34 min), 57% B (73 min), 100% B (90-98 min), 3% B (99-106 min). The PDA detector signal (250-380 nm) was recorded to monitor separation of peaks. Negative electrospray ionization mode was used for the MS detector by allowing 0.6 mL/min flow to enter the MS ion source using a split valve. Polyphenols were identified with the Information Dependent Analysis (IDA) scanning function of the mass spectrometer using a survey (Q1) scan between 150-1300 m/z and respective dependent (Q3) product ion scans between 80-1300 m/z. Ion source settings were as follows: spray voltage: -4500 V, source temperature: 500°C, curtain gas (N<sub>2</sub>) pressure: 40 psi, spray gas (N<sub>2</sub>) pressure: 30 psi, drying gas (N<sub>2</sub>) pressure: 30 psi. Chromatographic data were acquired and evaluated using the Analyst 1.6.3 software. Mass

spectra evaluation and compound identification was achieved using the RIKEN tandem mass spectral database (Sawada et al. 2012), via the scientific data found in the literature and by the use of fragmentation rules (McLafferty – Tureček 1993).

## 2.5 Statistics

In order to compare the respective antioxidant capacities of the extracts, ANOVA analysis was run using Statistica 11 (StatSoft Inc., Tulsa, USA) software with the Tukey HSD method.

## 3 RESULTS AND DISCUSSION

### 3.1 Evaluation of the TPC, FRAP and DPPH results

Table 1 includes the TPC, FRAP, and DPPH data of the samples indicating statistical comparison (ANOVA) for the 10 best results within each method. In all of the investigated taxa, the highest TPC was measured in green cone samples, followed by mature and opened cone samples. Overall the highest TPC was determined in the green cones of eastern hemlock ( $157.25 \pm 9.98$  mg GAE/g dw.), Lawson cypress ( $131.68 \pm 4.35$  mg GAE/g dw.), Japanese cedar ( $131.74 \pm 3.00$  mg GAE/g dw.) and dawn redwood ( $113.60 \pm 4.81$  mg GAE/g dw.). Respecting mature and opened cones highest TPC values were determined for dawn redwood (mature:  $91.25 \pm 3.69$  mg GAE/g dw., opened:  $60.16 \pm 8.23$  mg GAE/g dw.), Chinese arborvitae (mature:  $81.22 \pm 5.30$  mg GAE/g dw., opened:  $68.88 \pm 4.91$  mg GAE/g dw.), Japanese cedar (mature:  $74.18 \pm 2.09$  mg GAE/g dw., opened:  $57.41 \pm 2.93$  mg GAE/g dw.) and Norway spruce (mature:  $64.64 \pm 2.68$  mg GAE/g dw., opened:  $46.39 \pm 3.54$  mg GAE/g dw.).

According to literature data, Hofmann et al. (2020) determined high TPC levels for Norway spruce and eastern hemlock green cone samples. Horiba et al. (2016) found  $84.9 \pm 3.3$  mg GAE/g dw TPC in Japanese cedar cones (without seeds), which is comparable to the present results.

The overall highest TPC, determined for eastern hemlock green cones ( $157.25 \pm 9.98$  mg GAE/g dw.) was surprisingly higher than that of the related taxon, western hemlock ( $89.16 \pm 5.51$  mg GAE/g dw.). In fact, Hernes – Hedges (2004) reported the tannin content of western hemlock cones to be 3.13 wt.%; however, the authors did not document either the phenophase of cone maturity or the month of the sample collection. Hernes – Hedges (2004) also found that the bark and green needles contained more tannins compared to cones, yet did not investigate the amount of other types of polyphenols.

The limitation of the Folin-Ciocalteu assay (Singleton – Rossi 1965) is that it is known to interfere with other types of antioxidants (Prior et al. 2005, Everette et al. 2010). In fact, the TPC method is considered one of the >100 different assays currently used for the determination of antioxidant capacity and radical scavenging ability (Cornelli 2009). None of these assays is individually able to measure the total antioxidant power of all compounds in plant extracts. Therefore, the use of multiple assays to estimate the “overall” antioxidant potential of complex extracts is recommended (Ghiselli et al. 2000). The present study used the FRAP and the DPPH methods to provide further results on the antioxidant power of the samples.

Table 1. TPC<sup>1</sup>, FRAP<sup>2</sup>, and DPPH<sup>3</sup> antioxidant capacity of the cones (mean ± standard deviation). Different superscript letters indicate significant differences at  $p < 0.05$  (TPC, FRAP, DPPH) between the samples with the 10 best values

	TPC (mg GAE/g dw.)			FRAP (mg AAE/g dw.)			DPPH IC <sub>50</sub> (µg extractives/ml)		
	Green	Mature	Opened	Green	Mature	Opened	Green	Mature	Opened
<b>Atlas cedar</b>	88.41 ± 1.68	14.96 ± 2.24	7.46 ± 0.26	62.08 ± 3.13 <sup>a</sup>	4.48 ± 0.11	3.37 ± 0.10	21.44 ± 2.94	88.82 ± 12.86	56.92 ± 15.87
<b>European larch</b>	83.44 ± 4.27	25.98 ± 0.94	17.60 ± 2.15	55.96 ± 0.93	14.18 ± 0.83	4.09 ± 0.17	9.07 ± 1.39	12.53 ± 0.38	28.21 ± 6.84
<b>Norway spruce</b>	105.58 ± 7.92 <sup>ab</sup>	64.64 ± 2.68	46.39 ± 3.54	72.02 ± 8.76 <sup>ab</sup>	50.19 ± 2.08	28.35 ± 3.37	10.75 ± 0.32	9.38 ± 1.14	8.57 ± 0.17 <sup>ab</sup>
<b>Mountain pine</b>	95.76 ± 9.48 <sup>a</sup>	22.33 ± 3.31	15.96 ± 1.10	60.06 ± 2.77	9.34 ± 0.07	7.25 ± 0.19	7.87 ± 0.31 <sup>abc</sup>	27.83 ± 3.73	18.86 ± 0.14
<b>Black pine</b>	89.22 ± 4.79	19.70 ± 3.36	7.08 ± 0.34	58.21 ± 2.34	9.55 ± 0.52	4.50 ± 0.17	15.33 ± 1.39	45.90 ± 2.69	62.32 ± 1.90
<b>Scots pine</b>	46.30 ± 1.81	18.99 ± 1.44	13.19 ± 1.53	33.42 ± 3.12	9.41 ± 0.32	7.26 ± 0.14	72.40 ± 21.26	29.32 ± 1.10	22.88 ± 0.54
<b>Himalayan pine</b>	62.52 ± 5.09	17.76 ± 1.35	8.18 ± 0.97	38.84 ± 0.69	8.33 ± 0.56	3.85 ± 0.21	25.72 ± 3.50	54.76 ± 14.54	72.58 ± 7.23
<b>Eastern hemlock</b>	157.25 ± 9.98 <sup>d</sup>	56.13 ± 4.07	10.57 ± 1.69	100.11 ± 0.40 <sup>e</sup>	46.57 ± 1.02	5.94 ± 0.25	7.83 ± 0.29 <sup>abc</sup>	11.37 ± 0.67	17.74 ± 1.01
<b>Western hemlock</b>	89.16 ± 5.51	30.77 ± 2.22	10.01 ± 1.77	59.11 ± 1.73	31.03 ± 1.55	4.53 ± 0.09	11.16 ± 1.37	15.52 ± 0.84	40.44 ± 17.94
<b>Lawson cypress</b>	131.68 ± 4.35 <sup>c</sup>	20.61 ± 2.27	16.21 ± 2.11	89.42 ± 6.82 <sup>cde</sup>	9.18 ± 0.12	8.36 ± 0.13	7.23 ± 0.41 <sup>bc</sup>	22.46 ± 1.72	30.50 ± 6.72
<b>Bald cypress</b>	70.99 ± 4.49	52.20 ± 1.86	29.53 ± 3.96	57.34 ± 1.28	49.69 ± 5.07	42.42 ± 3.29	8.45 ± 0.74 <sup>ab</sup>	13.17 ± 2.13	13.42 ± 0.60
<b>Northern white-cedar</b>	93.71 ± 5.47 <sup>a</sup>	39.96 ± 2.59	31.38 ± 2.57	76.46 ± 3.44 <sup>abc</sup>	49.81 ± 0.11	18.54 ± 0.83	9.93 ± 0.62	9.21 ± 0.30	8.13 ± 0.55 <sup>ab</sup>
<b>Dawn redwood</b>	113.60 ± 4.81 <sup>b</sup>	91.25 ± 3.69 <sup>a</sup>	60.16 ± 8.23	129.16 ± 3.01 <sup>f</sup>	147.00 ± 6.83 <sup>g</sup>	61.43 ± 3.51	6.22 ± 0.42 <sup>c</sup>	4.42 ± 0.07 <sup>d</sup>	7.15 ± 0.87 <sup>bc</sup>
<b>Chinese arborvitae</b>	106.67 ± 2.76 <sup>ab</sup>	81.22 ± 5.30	68.88 ± 4.91	78.49 ± 1.55 <sup>bcd</sup>	93.12 ± 4.84 <sup>de</sup>	31.60 ± 2.02	9.56 ± 0.50	15.76 ± 0.45	17.27 ± 7.71
<b>Japanese cedar</b>	131.74 ± 3.00 <sup>c</sup>	74.18 ± 2.09	57.41 ± 2.93	60.87 ± 5.21	41.04 ± 2.08	24.16 ± 0.86	10.13 ± 0.76	10.55 ± 1.40	17.51 ± 0.56
<b>China fir</b>	92.24 ± 1.57 <sup>a</sup>	36.36 ± 2.29	35.94 ± 1.33	67.99 ± 8.88 <sup>ab</sup>	37.20 ± 2.68	20.65 ± 1.44	9.03 ± 1.19 <sup>a</sup>	13.79 ± 0.46	11.14 ± 0.45

1: Total polyphenol content

2: Ferric reducing antioxidant power

3: 2,2-diphenyl-1-picrylhydrazyl

Regarding FRAP results, green cone samples showed the best results in general. The only opposite tendency was observed with dawn redwood and Chinese arborvitae, where mature cones (D.r.:  $147.00 \pm 6.83$  mg AAE/g dw., C.a:  $93.12 \pm 4.84$  mg AAE/g dw.) had superior FRAP values compared to green cone results (D.r.:  $129.16 \pm 3.01$  mg AAE/g dw., C.a:  $78.49 \pm 1.55$  mg AAE/g dw.) showing excellent FRAP. Overall the best FRAP was determined for the green cones and opened cones of previous two taxa and for the green cones of eastern hemlock ( $100.11 \pm 0.40$  mg AAE/g dw.). According to Lesjak et al. (2011, 2014), the FRAP of *Juniperus* spp. cones varies between  $3.61 \pm 0.03$  mg AAE/g dw. (*Juniperus macrocarpa* Sibth. et Sm.) to  $35.26 \pm 1.12$  mg AAE/g dw. (*Juniperus sibirica* Burgsdorf.), which also indicates that big differences between related taxa can exist, as is the case with eastern ( $100.11 \pm 0.40$  mg AAE/g dw.) and western hemlock ( $59.11 \pm 1.73$  mg AAE/g dw.) cones in the present study.

The DPPH radical scavenging activity was determined using the IC<sub>50</sub> value (50% inhibition concentration), with low IC<sub>50</sub> indicating high antioxidant capacities. The DPPH results also showed the general decreasing tendency of the order green > mature > opened cones within a given taxon. The best results were obtained for the mature ( $4.42 \pm 0.07$  µg/ml) and green ( $6.22 \pm 0.42$  µg/ml) cones of dawn redwood, and for green cones of Lawson cypress ( $7.23 \pm 0.41$  µg/ml) and eastern hemlock ( $7.83 \pm 0.29$  µg/ml). In fact, the excellent DPPH activity (Bajpai et al. 2009, 2017) and bioactivity (Bajpai et al. 2007, 2009, Yoon et al. 2011) of dawn redwood cone extracts has already been reported in scientific literature.

The TPC, FRAP, and DPPH data makes it apparent that all of the three assays indicated different orders for the best results, which was attributed to the different compositions of the extracts as well as to the different working principle and selectivity of the assays (Apak et al. 2007, Müller et al. 2011).

In order to obtain a comprehensive measure of the overall antioxidant efficiency of the cone extracts and to consider the different selectivity of methods, the summarized evaluation of results of the three different methods was implemented.

### 3.2 Combined evaluation of the TPC, FRAP and DPPH results

Combined evaluation of the TPC, FRAP and DPPH was achieved using a scoring system (Hofmann et al. 2020) with the following calculation: For the TPC and FRAP results, 0 points were assigned to the weakest values and 1 to the best values within each assay, using linear approximation for the other values. In the case of DPPH assay, opposite scoring was used (lowest IC<sub>50</sub> value, score: 1; the highest IC<sub>50</sub>, score: 0). The respective scores of TPC, FRAP and DPPH were then summarized for each sample to estimate the measure of the overall antioxidant efficiency (Table 2).

Regarding the sum of scores, the highest scores – those with the best overall antioxidant power – were determined in the green cones of eastern hemlock (2.63), dawn redwood (2.56), Lawson cypress (2.40), Japanese cedar (2.16), Chinese arborvitae (2.13) and Norway spruce (2.06) and for the mature cones of dawn redwood (2.56). Interestingly eastern hemlock contained much higher overall antioxidant power compared to related western hemlock for green, mature and opened cone samples, showing big differences between respective samples; this discrepancy requires further research to determine an explanation.

Of these taxa, the bioactivity, antioxidant activity, or uses of their cone extracts have already been reported in the literature for Lawson cypress (Smith et al., 2007, Kilinc et al. 2015), dawn redwood (Bajpai et al. 2007, 2009, 2014, 2017, Yoon et al. 2011), Japanese cedar (Horiba et al. 2016) and Chinese arborvitae (Yogesh – Ali 2014).

However, no data on the polyphenolic composition and bioactivity of Norway spruce and eastern hemlock cone extracts exists in the scientific literature. Norway spruce is one of the most widespread coniferous tree species in Europe, possessing significant ecological, industrial

and economic significance (Meloni et al. 2007, Lamedica et al. 2011). Eastern hemlock is an ecologically important foundation species in forests of eastern North America (Clark et al. 2012) with a natural range extending from northern Georgia and Alabama to southern Canada and westward into the central Great Lakes states (McWilliams – Schmidt 2000). Information on molecular cone extract composition will provide a basis for the future research on the role these compounds play in possible bioactivity effects. Hence, the remainder of this article will focus on the identification of cone extractives, especially polyphenolic compounds found in the green cone tissues of Norway spruce and eastern hemlock.

*Table 2. Normalized values (scores) of the TPC<sup>1</sup>, FRAP<sup>2</sup>, and DPPH IC<sub>50</sub><sup>3</sup> values and the sum of scores for each sample representing the combined antioxidant values.*

	TPC <sup>1</sup>			FRAP <sup>2</sup>			DPPH IC <sub>50</sub> <sup>3</sup>			Sum of scores		
	Gr.	Mat.	Op.	Gr.	Mat.	Op.	Gr.	Mat.	Op.	Gr.	Mat.	Op.
Atlas cedar	0.54	0.05	0.00	0.41	0.01	0.00	0.80	0.00	0.38	1.75	0.06	0.38
European larch	0.51	0.13	0.07	0.37	0.08	0.00	0.94	0.90	0.72	1.82	1.10	0.79
Norway spruce	0.66	0.38	0.26	0.48	0.33	0.17	0.93	0.94	0.95	2.06	1.65	1.39
Mountain pine	0.59	0.10	0.06	0.39	0.04	0.03	0.96	0.72	0.83	1.94	0.87	0.92
Black pine	0.55	0.08	0.00	0.38	0.04	0.01	0.87	0.51	0.31	1.80	0.64	0.32
Scots pine	0.26	0.08	0.04	0.21	0.04	0.03	0.19	0.71	0.78	0.66	0.83	0.85
Himalayan pine	0.37	0.07	0.01	0.25	0.03	0.00	0.75	0.40	0.19	1.36	0.51	0.20
Eastern hemlock	1.00	0.33	0.02	0.67	0.30	0.02	0.96	0.92	0.84	2.63	1.54	0.88
Lawson cypress	0.83	0.09	0.06	0.60	0.04	0.03	0.97	0.79	0.69	2.40	0.92	0.79
Bald cypress	0.43	0.30	0.15	0.38	0.32	0.27	0.95	0.90	0.89	1.75	1.52	1.31
Northern white-cedar	0.58	0.22	0.16	0.51	0.32	0.11	0.93	0.94	0.96	2.02	1.49	1.22
Dawn redwood	0.71	0.56	0.35	0.88	1.00	0.40	0.98	1.00	0.97	2.56	2.56	1.75
Chinese arborvitae	0.66	0.49	0.41	0.52	0.62	0.20	0.94	0.87	0.85	2.13	1.98	1.46
Japanese cedar	0.83	0.45	0.34	0.40	0.26	0.14	0.93	0.93	0.84	2.16	1.64	1.32
China fir	0.57	0.19	0.19	0.45	0.24	0.12	0.95	0.89	0.92	1.96	1.32	1.23
Western hemlock	0.55	0.16	0.02	0.39	0.19	0.01	0.92	0.87	0.57	1.85	1.22	0.60

1: Total polyphenol content

2: Ferric reducing antioxidant power

3: 2,2-diphenyl-1-picrylhydrazyl

Gr.: green cones, Mat.: mature cones, Op.: opened cones

### 3.3 HPLC-MS/MS analyses

The identification of the molecular structure of the extractives in the cone extract solutions of Norway spruce and eastern hemlock has been accomplished using high-performance liquid chromatography/tandem mass spectrometry. *Figure 1.* depicts the HPLC chromatograms and *Table 3* includes the major compounds found in the extracts.

Altogether 82 compounds have been described and tentatively identified by tandem mass spectrometric fragmentation (MS/MS) data. The composition of the green cones of the two taxa is different, with both including low amounts of (+)-catechin (3), (-)-epicatechin (7), and procyanidin B dimers (1, 2, 4). Extracts included a large number of coumaric acid derivatives and flavonoid glycosides, yet not all of the compounds were found in both samples.



Quercetin-*O*-hexosides (18, 19) and taxifolin-*O*-hexosides (12,13) were found in both taxa; however, the pentose conjugate of quercetin (21) was only indicated in eastern hemlock. Interestingly, isorhamnetin-*O*-hexosides (27, 28) were only found in Norway spruce. The most abundant class of flavonoid conjugates were the kaempferol derivatives (mostly glycosides) with a total count of 10 compounds. Out of these compounds only kaempferol-*O*-hexoside (25), kaempferol-*O*-ruinoside (37) and kaempferol-rhamnose-hexose-rhamnose (50) were detected in the green cones of both taxa. The *O*-rutinoside (24), *O*-pentoside (29, 30, 31), *O*-rhamnoside (33), acetyl-hexoside (34), and an unknown derivative (46) of kaempferol were exclusively detected in eastern hemlock. Regarding flavonoid glycosides, the presence of acylated kaempferol conjugates (e.g. 34) are especially interesting as these types of compounds were shown to have excellent antioxidant properties and to contribute significantly to antibacterial effects of plant extracts (Mellou et al. 2005), which highlights the importance in finding matrices with high content of acylated flavonols (García-Villalba et al. 2017).

The presence of coumaric acid as part of the compounds was evidenced by the simultaneous presence of the 163, 145, and 119 *m/z* ions in the MS/MS spectra of the compounds corresponding to the  $[M-H]^-$ ,  $[M-H_2O-H]^-$  and  $[M-CO_2-H]^-$  fragment ions, with M representing coumaric acid molecule. The structure of coumaric acid derivatives are often left unidentified using ion trap or triple quadrupole mass spectrometers (Spínola et al. 2016, Llorent-Martínez et al. 2019), as the MS<sup>n</sup> mass spectra are only suitable to indicate characteristic fragments and losses during the fragmentation process of the molecules, justifying the simultaneous presence of the coumaric acid fragments at 119, 145 and 163 *m/z*. A more precise and informative analysis of the structure of these compounds could be conducted in the future with the use of TOF (time-of-flight) mass spectrometry by determining accurate mass of the compounds (Vilhena et al. 2020). Coumaric acid derivatives 47, 48, 49, 59, and 66 were only indicated in Norway spruce, while compounds 55, 60, and 65 were found exclusively in eastern hemlock and compound 51 in the green cone extracts of both taxa.

Piceatannol isomers (15, 16) and their *O*-hexoside conjugates (possibly astringin isomers, 10, 11) were evidenced from spruce samples only.

Chlorogenic acid isomers (5, 6) were only found in eastern hemlock. Other compounds were left unidentified only with MS/MS data for future identification of their structure.

According to *Table 3* and comparing peak heights in *Figure 1*, the most abundant compounds in the green cone extract of Norway spruce were astringin isomers (10, 11), unidentified compounds 8, 58, 68, 69, 70 and coumaric acid derivative 51, while in eastern hemlock they were chlorogenic acid isomers 5, 6, kaempferol-rhamnose-hexose-rhamnose 50, and unidentified compounds 68, 69, 70, and 79.

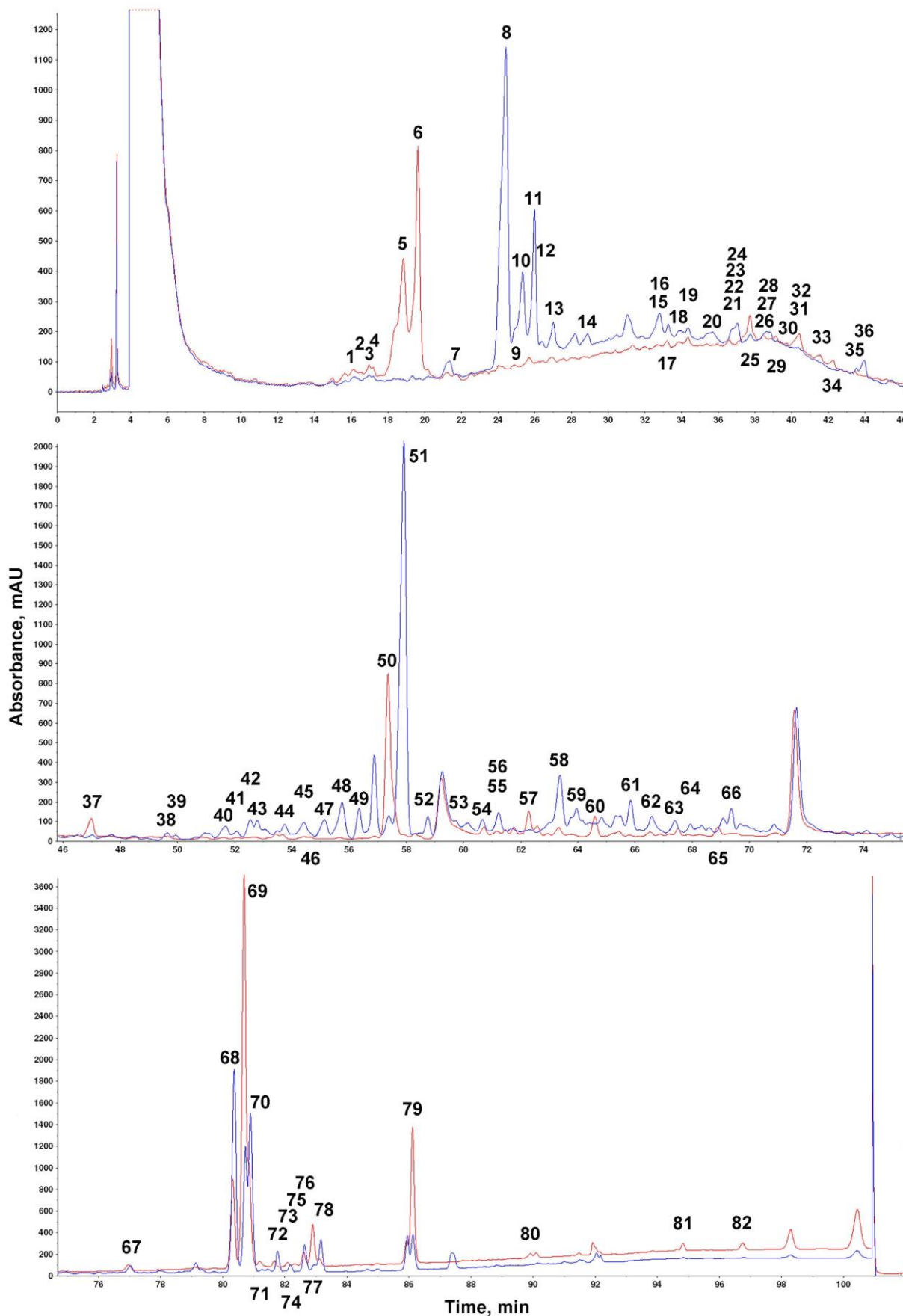


Figure 1. The PDA (250-380 nm) chromatogram of the green cone extracts of Norway spruce (blue) and eastern hemlock (red).

Table 3. Tentative chromatographic/mass spectrometric identification of the polyphenols in the green cones of Norway spruce (S) and eastern hemlock (H)

Peak	$t_r$ (min)	Compound	S	H	[M-H] <sup>-</sup> m/z	MS/MS m/z
1	15.8	Procyanidin B dimer	x	x	577	425, 407, 289, 245, 125
2	16.2	Procyanidin B dimer	x	x	577	425, 407, 289, 245, 125
3	17.0	(+)-Catechin	x	x	289	245, 203, 125, 123, 109
4	17.2	Procyanidin B dimer	x	x	577	425, 407, 289, 245, 125
5	18.9	Chlorogenic acid isomer		x	353	191, 179, 161, 135
6	19.7	Chlorogenic acid isomer		x	353	191, 179, 161, 135
7	21.7	(-)-Epicatechin	x	x	289	245, 203, 125, 123, 109
8	24.0	Unidentified	x		no ion	no negative ions
9	25.0	Unidentified	x		no ion	no negative ions
10	25.3	Piceatannol- <i>O</i> -hexoside (astringin)	x		405	243, 225, 201
11	26.0	Piceatannol- <i>O</i> -hexoside (astringin)	x		405	243, 225, 201
12	26.3	Taxifolin- <i>O</i> -hexoside	x	x	465	447, 437, 303, 285, 259, 217, 179, 125
13	27.1	Taxifolin- <i>O</i> -hexoside	x	x	465	447, 437, 303, 285, 259, 217, 179, 125
14	29.0	Unidentified	x		285	241, 217, 199
15	32.6	Piceatannol	x		243	225, 201, 175, 174
16	32.8	Piceatannol	x		243	225, 201, 175, 174
17	33.3	Unidentified	x		257	241, 211,
18	33.9	Quercetin- <i>O</i> -hexoside	x	x	463	301, 300, 271, 255, 179
19	34.4	Quercetin- <i>O</i> -hexoside	x	x	463	301, 300, 271, 255, 179
20	35.4	Unidentified	x		359	341, 311, 297, 282, 195, 163, 145
21	36.6	Quercetin- <i>O</i> -pentoside		x	433	301, 300, 271, 255, 243, 179
22	36.8	Unidentified	x		373	358, 313, 305
23	37.0	Unidentified	x		359	341, 311, 297, 282, 195, 163, 145
24	37.2	Kaempferol- <i>O</i> -rutinoside		x	593	447, 285, 284, 255, 227
25	37.7	Kaempferol- <i>O</i> -hexoside	x	x	447	285, 284, 255, 227
26	38.2	Unidentified- <i>O</i> -hexoside		x	431	268, 269
27	38.6	Isorhamnetin- <i>O</i> -hexoside	x		477	315, 314, 300, 299, 271
28	38.9	Isorhamnetin- <i>O</i> -hexoside	x		477	315, 314, 300, 299, 271
29	39.2	Kaempferol- <i>O</i> -pentoside		x	417	285, 284, 255, 227
30	39.8	Kaempferol- <i>O</i> -pentoside		x	417	285, 284, 255, 227
31	40.4	Kaempferol- <i>O</i> -pentoside		x	417	285, 284, 255, 227
32	40.5	Unidentified- <i>O</i> -hexoside	x	x	447	315, 285, 217, 199
33	41.6	Kaempferol- <i>O</i> -rhamnoside		x	431	285, 284, 255, 277
34	42.2	Kaempferol-acetyl-hexoside		x	489	429, 285, 284, 255, 227
35	43.6	Unidentified	x	x	351	333, 315, 275, 251
36	43.9	Unidentified	x		291	245, 175
37	47.0	Kaempferol- <i>O</i> -rutinoside	x	x	593	447, 285, 284, 255, 227
38	49.8	Unidentified	x	x	351	333, 315, 275, 251
39	50.0	Unidentified	x		367	349, 321, 247
40	51.7	Unidentified	x		377	331
41	52.0	Unidentified	x		331	313, 273, 241, 185
42	52.6	Unidentified	x		349	331, 287, 251, 244, 207, 189, 163
43	52.8	Unidentified	x		405	375, 337, 327, 275
44	53.7	Unidentified	x		401	333, 315, 257
45	54.4	Unidentified	x		521	179, 162, 146, 135
46	54.7	Kaempferol derivative		x	635	285, 284
47	55.1	Coumaric acid derivative	x		445	427, 397, 349, 277, 251, 163, 145, 119
48	55.8	Coumaric acid derivative	x		475	457, 427, 281, 163, 145, 119
49	56.4	Coumaric acid derivative	x		505	487, 457, 311, 163, 145, 119
50	57.4	Kaempferol-rhamn.-hex.-rhamn.	x	x	739	593, 453, 285, 284, 255, 229

Table 3 cont. Tentative chromatographic/mass spectrometric identification of the polyphenols in the green cones of Norway spruce (S) and eastern hemlock (H)

Peak	$t_r$ (min)	Compound	S	H	[M-H] <sup>-</sup> m/z	MS/MS m/z
51	58.0	Coumaric acid derivative	x	x	505	491, 477, 342, 327, 312, 177, 163, 119
52	58.8	Unidentified	x		535	520, 491, 341, 326, 193, 179, 134
53	59.7	Unidentified	x	x	445	417, 399, 315
54	60.7	Unidentified	x	x	401	333, 315, 289, 245
55	61.1	Coumaric acid derivative		x	549	489, 353, 311, 163, 145, 119
56	61.2	Unidentified	x		349	331, 289, 245
57	62.1	Unidentified	x	x	399	367, 331, 299
58	63.4	Unidentified	x	x	385	317, 299, 253
59	64.0	Coumaric acid derivative	x		667	521, 403, 323, 163, 145, 119
60	64.6	Coumaric acid derivative		x	653	638, 507, 489, 353, 329, 177, 163, 145, 119
61	66.0	Unidentified	x		383	355, 315, 297
62	66.6	Unidentified	x		383	315, 299, 269
63	67.4	Unidentified	x		471	425, 403, 353, 325, 285
64	68.0	Unidentified	x	x	381	313, 269
65	68.9	Coumaric acid derivative		x	651	487, 472, 341, 326, 266, 163, 145, 119
66	69.4	Coumaric acid derivative	x		649	441, 426, 411, 321, 291, 253, 163, 145, 119
67	77.0	Unidentified	x	x	429	381, 299, 265
68	80.4	Unidentified	x	x	687	657, 301
69	80.7	Unidentified	x	x	397	301
70	80.9	Unidentified	x	x	431	401, 383, 301
71	81.2	Unidentified		x	469	425, 410, 384, 367, 339, 285
72	81.7	Unidentified	x		455	409, 391, 387, 355, 287
73	82.1	Unidentified		x	957	467, 423, 381
74	82.2	Unidentified	x		455	409, 391, 387, 355, 287
75	82.4	Unidentified		x	935	467, 424, 382, 265
76	82.6	Unidentified	x	x	721	417, 335, 317
77	82.9	Unidentified		x	467	449, 423, 408, 382, 338
78	83.1	Unidentified	x	x	633	333, 317, 315, 299
79	86.1	Unidentified		x	635	591, 333, 317, 301, 271
80	89.9	Unidentified		x	769	725, 467, 301
81	94.8	Unidentified		x	501	486
82	96.7	Unidentified		x	529	514

rhamn.: rhamnose; hex.,: hexose

#### 4 CONCLUSIONS

The present study compared and evaluated the antioxidant capacity of the cone extracts of 16 selected coniferous taxa. The overall antioxidant power was determined by a scoring system that combined the results of the three antioxidant assays used in the study. The best antioxidant properties were determined for green cones, followed by mature and opened cones for each taxon. The highest scores were found for *Tsuga canadensis*, *Metasequoia glyptostroboides*, *Chamaecyparis lawsoniana*, *Cryptomeria japonica*, *Thuja orientalis* and *Picea abies*, which contained high amounts of antioxidants in both green and mature cones. The high-performance liquid chromatographic/tandem mass spectrometric profiling of the green cone extractives of *Picea abies* and *Tsuga canadensis* was carried out and overall 82 compounds have been tentatively identified from these samples for the first time, including kaempferol-, taxifolin-, quercetin- and isorhamnetin-*O*-glycosides, coumaric acid derivatives, chlorogenic acids,

piceatannol and its conjugates, and flavan-3-ol compounds. Presented chromatographic/mass spectrometric data on the polyphenolic composition of the green cone extracts contributes to the determination of the structure of unidentified compounds and to the research on the role of extractives in determining the bioactivity of cone extracts. To enhance practical use of this study's results, future research will focus on the antibacterial and antifungal properties of the investigated cone extracts with the highest antioxidant capacity.

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